

## EcoRI

5'-G A A T T C-3'  
3'-C T T A A G-5'

Cat. No.	Size
E2150-01	10 000 units
E2150-02	50 000 units
E2150-03	250 000 units

**Reaction Temperature:** 37°C

**Inactivation Temperature (20 min):** 65°C

**Prototype:** EcoRI

**Source:** *Escherichia coli* RY 13

### Package Contents:

- **EcoRI**
- **10x Reaction Buffer High**
- **BSA with Triton X-100 [100x]**  
Added as separate component to prevent reaction buffer precipitation.
- **Dilution Buffer # 3**  
Added for enzymes exceeding 10 U/μl in concentration. High protein concentration warrants optimal stability during prolonged storage. Use dilution buffer to prepare short term working stocks (5-10 U/μl, non-freezing at -20°C).

**Storage Conditions:** Store at -20°C

### Double Digestion – Buffer Compatibility:

Buffer	% Rel. Activity
<b>Low</b>	25*
<b>Medium</b>	100*#
<b>High</b>	<b>100</b> #
<b>Acet</b>	50

\* - The enzyme exhibits star activity under certain conditions.

# - **Adding 0,025% Triton-X100 and BSA to reaction is required for optimal digestion.**

### Recommended Buffer: High

(or compatible third party buffers)

### Restriction Enzyme Buffer Compatibility:

Both, enzyme and buffers are fully compatible to restrictases and buffer systems from other manufacturers and can be used along in double digestions. To obtain best results, consult the corresponding manuals of all involved products.

### DNA Methylation:

No inhibition: dam, dcm, EcoKI

Potential inhibition : CpG

### Standard Reaction Protocol:

**Mix** the following reaction components:

1-2 μg pure DNA or 10 μl PCR product (=~0.1-2 μg DNA)

5 μl 10x Buffer High

0.5 μl BSA with Triton X-100[100x]

1-2 U EcoRI (use 1 U / μg DNA, < 10 % React. Volume!)

*Tips:* Add enzyme as last component. Mix components well before adding enzyme. After enzyme addition, mix gently by pipetting. Do not vortex. High (excess) amounts of enzyme can greatly speed up the reaction.

@ 50 μl H<sub>2</sub>O, nuclease free

**Incubate** for 1 h at 37°C

To obtain complete digestion of high molecular weight DNA, (e.g. plant genomic DNA), add excess amounts of enzyme and prolong the incubation time.

**Stop** reaction by alternatively

(a) Addition of 2.1 μl EDTA pH 8.0 [0.5 M], final 20 mM or

(b) Heat Inactivation

20 min at 65°C or

(c) Spin Column DNA Purification

(e.g. EURx PCR/DNA CleanUp Kit, Cat.No. E3520) or

(d) Gel Electrophoresis and Single Band Excision

(e.g. EURx AgaroseOut DNA Kit, Cat.No. E3540) or

(e) Phenol-Chloroform Extraction or Ethanol Precipitation.

### Non-optimal buffer conditions:

To compensate for the lack of enzyme activity, increase the amount of enzyme and / or reaction time accordingly. The following values may serve as orientation:

1. **Enzyme amount:** Instead of 1 U enzyme, use ~4 U of enzyme in buffers providing 25 % rel. activity, ~2 U in 50 %, ~1.5 U in 75 % or ~1 U in 100 %, respectively.
2. **Reaction time:** Increase by ~1.3-fold (75 % rel. activity), ~2-fold (50 %) or ~4-fold (25 %).

### Unit Definition:

One unit is the amount of enzyme required to completely digest 1 μg of Lambda DNA in 1 hr in a total reaction volume of 50 μl. Enzyme activity was determined in the recommended reaction buffer.

### Reaction Buffer:

**1 x High Buffer:** 50 mM Tris-HCl (pH 7.5 at 37°C), 10 mM MgCl<sub>2</sub>, 100 mM NaCl, 1 mM dithiothreitol.

To be supplemented with 100 μg/ml bovine serum albumin and 0,025% Triton X-100.

### Storage Buffer:

10 mM potassium phosphate (pH 7.5), 300 mM NaCl, 0.1 mM EDTA, 1 mM dithiothreitol, 0.2 % (v/v) Triton™X-100, 100 μg/ml bovine serum albumin, 50 % (v/v) glycerol.

### Quality Control:

All preparations are assayed for contaminating endonuclease, 3'-exonuclease, 5'-exonuclease/5'-phosphatase, as well as non-specific single- and double-stranded DNase activities.